## SHORT COMMUNICATION

# CHEMICAL INDUCTION OF LOW TEMPERATURE BREAKDOWN IN APPLES

R. B. H. WILLS

C.S.I.R.O. Division of Food Preservation, Ryde, N.S.W., Australia

and

K. J. SCOTT

N.S.W. Department of Agriculture, located at C.S.I.R.O., Ryde, N.S.W., Australia

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Abstract—Jonathan apples injected with acetate, mevalonate, malonate, formate, butyrate, benzoate, caffeine, amytal, adenosine triphosphate and octanol showed an increase in low temperature breakdown after storage at  $-1^{\circ}$ . Injection of calcium resulted in a decrease in the incidence of breakdown. It is suggested that isoprenoid metabolism may be involved in the metabolic events leading to breakdown.

#### INTRODUCTION

When stored below 5° certain varieties of apples are susceptible to the physiological disorder, low temperature breakdown. The disorder, which appears initially as browning in the outer cortex, has long been considered to result from an alteration in the metabolism of the fruit at low temperature.1

Wills<sup>2</sup> examined the volatiles given off by apples and found that, when the loss of certain acetate esters increased, the susceptibility of the fruit to breakdown decreased. He suggested that accumulation of acetate in the fruit could be involved in breakdown. Wills, Scott and McGlasson<sup>3</sup> examined the role of acetate and found that, early in storage, acetic acid accumulated to higher levels in fruit that were more susceptible to breakdown but later fell to low levels in all fruit before the appearance of visible symptoms of breakdown. They were able to induce the disorder by injecting acetic acid into the core of fruit but it appeared only after storage for many weeks. It was concluded that acetate itself was not causing breakdown but that the disorder was a result of its effect on the metabolism of the stored fruit.

In this paper we have attempted to determine which aspect of acetate metabolism might be involved in breakdown, by injecting a large number of substances that can be formed from acetate. We have also injected the major volatile compounds of apples, other compounds which have been associated with breakdown, namely oxalacetate,4 acetaldehyde,<sup>5</sup> sorbitol,<sup>6</sup> and compounds containing nitrogen and phosphorus,<sup>7</sup> and salts of several

<sup>&</sup>lt;sup>1</sup> F. Kidd and C. West, Gt Brit. Rep. Food Invest. Bd. 42 (1927).

<sup>&</sup>lt;sup>2</sup> R. B. H. WILLS, J. Sci. Food Agri. 19, 354 (1968)

<sup>&</sup>lt;sup>3</sup> R. B. H. WILLS, K. J. SCOTT and W. B. McGLASSON, J. Sci. Food Agri. 21, 42 (1970).

<sup>&</sup>lt;sup>4</sup> A. C. HULME, W. H. SMITH and L. S. C. WOOLTORTON, J. Sci. Food Agri. 15, 303 (1964). <sup>5</sup> H. CLUSTERS, Physiol. Plantarum 18, 85 (1965).

<sup>&</sup>lt;sup>6</sup> J. C. Fidler and C. J. North, J. Hort. Sci. 43, 429 (1968).

L. W. TILLER, H. S. ROBERTS and E. G. BOLLARD, New Zealand Dep. Sci. Ind. Res. Bull. No. 129 (1959).

mineral elements present in the fruit. The fruits were stored for 1 week at  $-1^{\circ}$ , and each injected with 80  $\mu$ mole of the substance used and then stored at  $-1^{\circ}$  for 3–9 months before being inspected for breakdown.

Table 1. Incidence of Breakdown in apples injected with various chemicals

Compounds affecting breakdown	% Bd.	Sig. diff.	Comps. not diff. from control	% Bđ.	Solvent control	% Bd.
Expt. 1						
R-Mevalonate	59	. l. 1 1	Duramete	42	Water	20
Malonate	53	+++	Pyruvate L-Malate	42 36	Water	30
Formate	49	++	Acetaldehyde	36		
Acetate	48	÷÷+	cis-Oxalacetate	35		
Butyrate	47	+-	L-Serine	34		
•		'	L-Glutamate	34		
			Propionate	32		
			Acetoacetate	31		
			D-3-Phosphoglycerate	24		
			Isopentyl acetate	45	Ethanol	33
			Isopentanol	40		
			Butyl acetate	38		
			Hexyl acetate	37		
			Ethyl butyrate 32			
7t 2			Hexanol	31		
Expt. 2	60	1 1 1	and .	••		
Benzoate Caffeine	69	+++	Thiourea	30	Water	18
aneme Amytal	49	+++	α-D-Glucose-1-phosphate	26		
ingtal cetate	43 40	++	L-Phenylalanine	26		
A.T.P.	36	++	Sodium carbonate	24		
1. I . F .	30	+	Potassium carbonate Phosphoric acid	23		
			Shikimate	23		
Expt. 3			Snikimate	19		
Acetate	61	++	Sorbitol	43	Water	41
expt. 4						
Octanol	52	+++	Octyl acetate	24	Ethanol	12
			Hexanoic acid	23	Emanor	14
			Ethyl hexanoate	21	<del>-</del>	
			Hexanol 16			
			Heptanol	16		
			Hexyl butyrate	14		
			Hexyl acetate	xyl acetate 14		
Expt. 5			Hexanol	5		
cetate	59		Hydrophlaria asid	10	117-4	
Calcium chloride	39 24	+	Hydrochloric acid	48	Water	42
aloidin chionide	<b>∠</b> →		Ammonium chloride	46		
			Magnesium chloride Potassium chloride	40		
			rotassium entorige	39		

<sup>(+++)</sup>, (++) And (+) indicate that the compound produced significantly more breakdown than control fruit at 0·1%, 1% and 5% levels respectively and (-) indicates less breakdown than control at 5% level.

## RESULTS AND DISCUSSION

The results (Table 1) show that a number of substances were effective in increasing the the incidence of breakdown. Mevalonate was the most effective of the substances associated with acetate metabolism. Malonate was also effective, but a number of other compounds had no significant effect under the conditions used. Both acetate and mevalonate are precursors for isoprenoid synthesis and the greater activity of mevalonate perhaps points to isoprenoid involvement in breakdown. Octanol, which was effective at the same level as mevalonate, might affect isoprenoid metabolism as could malonate. Octanol has been shown to behave like a monoterpene in its feedback inhibition of prenyl transferase<sup>8</sup> and malonate was found to be an intermediate in the synthesis of mevalonate in animal systems.<sup>9</sup>

No explanation can be advanced for the activity of other substances that increased breakdown, though the effect of caffeine, amytal and adenosine may be due to a requirement for nitrogen in breakdown processes in view of the increase in breakdown observed following high nitrogen levels of fertilization of the tree during growth.<sup>7</sup> The effect of calcium in reducing breakdown confirms the work of Perring<sup>10</sup> who found lower levels of calcium in fruit susceptible to breakdown, and Rasmussen,<sup>11</sup> who reduced breakdown by spraying with calcium during growth.

Work is proceeding in this laboratory to determine the importance of isoprenoid metabolism, and also purines and benzoate, in the production of breakdown.

## **EXPERIMENTAL**

Fruit was harvested in 1969 and 1970 from commercial orchards in N.S.W., Australia, and randomly distributed into units, each of 25 fruits. Fruits were injected in the core areas with 80  $\mu$ mole of the free acids or pure compounds, dissolved in 0.2 ml of H<sub>2</sub>O or EtOH, except for D-3-phosphoglycerate, L-glutamate, benzoate, shikimate, L-phenylalanine and amytal which were injected as sodium salts, and acetoacetate and R-mevalonate which were injected as the methyl ester and as the RS-lactone respectively. As only the racemate of mevalonate was available, 160  $\mu$ mole of the mixture was injected into the fruit. The amount of ATP added was 80  $\mu$ mole with respect to the concentration of phosphorus, equivalent to 27  $\mu$ mole of ATP.

Expt.	Time at -1° (weeks)	No. of fruit per treatment	S.E. of mean (angles)	
1	13	200		
2	40	100	$\pm 3.4^{\circ}$ on 39 d.f.	
3	18	125	$\pm 2.0^{\circ}$ on 12 d.f.	
4	16	100	$\pm 4.1^{\circ}$ on 30 d.f.	
5	16	100	$\pm 3.3^{\circ}$ on 21 d.f.	

TABLE 2. STORAGE DATA OF APPLES INJECTED WITH VARIOUS CHEMICALS

After storage at  $-1^{\circ}$ , the fruits were kept at  $20^{\circ}$  for 7 days after which they were examined for the occurrence of breakdown. The percentages of fruit affected with breakdown were transformed to angles by the arcsin transformation and standard errors were calculated. The levels of statistical significance were determined by t-test. Table 2 gives the standard errors for the groups of injections and the time each group was stored at  $-1^{\circ}$ .

Acknowledgements—We thank Mr. W. Bailey for technical assistance.

<sup>&</sup>lt;sup>8</sup> G. POPJAK, P. W. HOLLOWAY, R. P. M. BOND and M. ROBERTS, Biochem. J. III, 333 (1969).

<sup>&</sup>lt;sup>9</sup> J. D. Brodie, G. Wasson and J. W. Porter, J. Biol. Chem. 238, 1294 (1963).

<sup>&</sup>lt;sup>10</sup> M. A. PERRING, J. Sci. Food Agri. 19, 186 (1968).

<sup>11</sup> M D DACISTICSEN Frhomefruotanleren 32, 283 (1965).